SYNTHESIS OF "BRANCHED" TRINUCLEOTIDE USING THE H-PHOSPHONATE CHEMISTRY

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Abstract: Highly electrophilic phosphorylating species, generated from an appropriately protected 5'-O-(1-H-phosphonate) nucleoside block 4 or 5 and pivaloyl chloride, followed by an oxidation step, has been used for the first time to introduce the branching phosphate vicinal to an internucleotidyl $2' \rightarrow 5'$ or $3' \rightarrow 5'$ diribonucleoside monophosphate function in 2 or 3, providing an alternative route to the synthesis of the lariat structure 1a.

The precise mechanism of excision of introns and ligation of exons from the pre-mRNA (splicing) is one of the most fundamental events in the living process of the eukaryotes. In certain types of splicing reactions, the linear $3' \rightarrow 5'$ phosphodiester linked mRNA of the intron is branched specifically at adenosine to form an additional $2' \rightarrow 5'$ phosphodiester linked circular mRNA intron called lariat^{1,2}. Since the isolation of the "branched" RNA, there have been several reports of the synthesis of the core "branched" triribonucleotides $A_{3'p5'}^{2'p5'G}$ 1 which particularly demonstrated the synthetic difficulties in introducing two phosphodiesters to the vicinal 2' and 3' hydroxyl groups at the branch-point adenosine 3^{-8} . It has thus emerged that, for the introduction of the second phosphate residue vicinal to the internucleotidic phosphate at the branch-point, the internucleotidic phosphate should be as poorly electrophilic as possible for it to be stable and resistant to the isomerization reactions due to the participation of the vicinal hydroxyl group 9-11. It is apparent that it is the internucleotidic *phosphodiester* function only which fulfils these criteria satisfactorily since building blocks such as 2 and 3 are clearly isolable in a stable and pure state^{5,6a,7}. The fulfilment of these criteria is essential for sequential, regiospecific introduction of dissimilar nucleotide residues in a high yield at the branch-point for the synthesis of lariat structures such as 1. While an internucleotidic phosphite-triester has been found to be stable enough in situ for the introduction of the second phosphite residue to the vicinal hydroxyl group in Ogilvie's approach³, but an internucleotidic phosphotriester^{9,10} or an H-phosphonate¹¹, vicinal to a hydroxyl group, are unstable due to the participation of the vicinal hydroxyl group. One other important observation is that stable building blocks such as 2 and 3 only react with the highly reactive electrophilic species generated by the reaction of an appropriately protected 5'-phosphoroamidite^{3,5-8}, such as 6, in presence of tetrazole at room temperature in order to introduce the second branching phosphate to give 1 upon deprotection. The only exception to this general observation has been reported by Hata and his coworkers who employed cyclohexylammonium S,S-diphenylphosphorothioate and mesitylene-2,4-disulfonyl chloride to introduce the second phosphate residue to the 3'-O-phosphorobisanilidate derivative⁴ which has been isolated as the isomerized product from the corresponding 2'-O-phosphorobisanilidate derivative.



Tol = p-toluoyl, Tbb = t-butylbenzoyl, Px = 9-phenylxanthenyl (pixyl), $U^{Bz} = N^3$ -benzoyluracil-1-yl $A^{Bz} = N^6$ -benzoyl-9-adeninyl, Ac = acetyl, A = 9-adeninyl, G = 9-guaninyl, U = 1-uracilyl, C = 1-cytosinyl $U^{NP} = O^4$ -(2-nitrophenyl)uracil-1-yl.

We herein report for the first time that the H-phosphonate chemistry 12,13 can be also efficiently used for the purpose of introduction of the second branching phosphate vicinal to the phosphodiester function in appropriately protected dimeric blocks such as 2 or 3, and thus provides a convenient alternative synthetic approach to the preparation of 1a. The apparent advantage of being able to use the 5'-O-(1-H-phosphonate) building blocks such as 4 or 5, compared to a 5'-O-phosphiteamidite block such as 6, is that they can be easily prepared and purified without any special care and are also stable over a prolonged period of storage12,13

Appropriately protected diribonucleoside $(2^{\prime} \rightarrow 5^{\prime})$ monophosphate 2^{6c} was thus charged with the Hphosphonate block 4^{14} (5 equiv.) in presence of a large excess of freshly distilled pivaloyl chloride (30) equiv.) in dry pyridine, added in two portions in ten minutes' interval at room temperature. The reaction was kept stirring for ca. 30 min when it was worked up, after an oxidation step, in the usual manner 12,13. The reaction products were then directly deprotected 6a by the treatment of aqueous ammonia at ~ 50 °C to give the branched trimer 1a¹⁵ in 34 % yield.

In order to show that the introduction of the second phosphate group to the 2'-hydroxyl function vicinal to a phosphodiester is equally facile, we have prepared diribonucleoside $(3^{-} \rightarrow 5^{-})$ monophosphate 7 in 68 % yield by a direct condensation of 8 with the H-phosphonate block 4^{14} (5 equiv.) in the presence of pivalov chloride (30 equiv.) in dry pyridine followed by an oxidation step 12,13. Such facile preparation of 7 circumvents the preparation of phosphate-protected triester precursor 9 which had to be necessarily prepared in earlier methods to give the diester 7 through the phosphate deprotection step⁵⁻⁷. The 2'-O-pixyl group was then removed from compound 7 by a brief acid treatment with trichloroacetic acid in 1% ethanol-CHCl3 mixture to give 3^{18} (62 %) which was condensed in the usual manner 12,13 with the H-phosphonate block 5^{16} and subsequently deprotected by aqueous ammonia to give the branched trimer $1a^{15}$ in 21 % yield essentially following a procedure described for the reaction of $2 + 4 \rightarrow 1a$. Incidentally, this synthesis of 1a from 8 [8 + 4 \rightarrow 7; 7 \rightarrow 3; 3 + 5 \rightarrow 1a] constitutes, to this date, its shortest synthesis in a high overall yield using the H-phosphonate methodology 12,13 entirely for the introduction of both of its vicinal phosphate functions.

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References

- T.R. Cech and B.L. Bass, Ann. Rev. Biochem. 55, 599-629 (1986). 1.
- R.A. Padgett, P.J. Grabowski, M.M. Konarska, S. Seiter and P.A. Sharp, Ann. Rev. Biochem. 55, 2. 1119-1150 (1981). M.J. Damha, R.T. Pon and K.K. Ogilvie, *Tetrahedron Lett.* 26, 4839 (1985).
- 3.
- M. Sekine and T. Hata, J. Am. Chem. Soc. 107, 5813 (1985); M. Sekine, J. Heikkilä and T. Hata. 4. Tetrahedron Lett., 28, 5691 (1987).
- R. Kierzek, D.W. Kopp, M. Edmonds and M.H. Caruthers, Nucleic Acids Res., 14, 4751 (1986). 5.
- (a) J.M. Vial, N. Balgobin, G. Remaud, A. Nyilas and J. Chattopadhyaya, *Nucleosides and Nucleotides*, 6, 209 (1987); (b) G. Remaud, X.-X. Zhou, B. Öberg and J. Chattopadhyaya, in "Reviews of heteroatom Chemistry", vol. 1, ed. S. Oae, MYU publishing Inc., Tokyo, 1988, p. 340-366; (c) X.-X. Zhou, A. Nyilas, G. Remaud and J. Chattopadhyaya, *Tetrahedron*, 43, 4685 6.

(1987); (d) X.-X. Zhou, G. Remaud and J. Chattopadhyaya, *Tetrahedron*, 44, 6471 (1988); (e) N. Balgobin, A. Földesi, G. Remaud and J. Chattopadhyaya, *Tetrahedron* (in press, 1988).

- 7. S. Huss, G. Gosselin and J.-L. Imbach, J. Org. Chem., 53, 499 (1988).
- 8. J.L. Fourrey, J. Varenne, C. Fontaine, E. Guittet and Z.W. Yang, *Tetrahedron Lett.*, 28, 1769 (1987).
- 9. T. Pathak and J. Chattopadhyaya, Acta Chem. Scand., B39, 799 (1985).
- 10. C.B. Reese and P. Skone, Nucleic Acids Res., 13, 5215 (1985).
- 11. S. Huss, G. Gosselin, J. Stawinski, R. Strömberg and J.-L. Imbach, Nucleosides and Nucleotides, 7, 321 (1988).
- 12. P.J. Garegg, T. Regberg, J. Stawinski and R. Strömberg, Chem. Scripta, 25, 280 (1985).
- 13. B.C. Froehler and M.D. Matteucci, Tetrahedron Lett., 21, 469 (1986).
- 14. ¹H-NMR (90 MHz, CDCl₃ + TMS): 8.10 (d, 8.1 Hz, 1H, H-6); 8.0-7.4 (m, 5H, Bz); 6.99 (d, 626.0 Hz, 1H, HP); 6.21 (d, 5.6 Hz, 1H, H-1'); 5.55 (d, 8.1 Hz, 1H, H-5); 5.7-5.3 (m, 2H, H-2',3'); 4.5-3.9 (m, 3H, H-4',5'); 2.8 (m, 6H, CH_2 CH₃); 2.11 & 1.99 (2 x s, 6H, Ac); 1.5 (m, 9H, CH_3 CH₂). ³¹P-NMR (CDCl₃ + ext. standard 85% aq. H₃PO₄): +3.49 (dt, ¹J_{H,P} = 626.0 Hz, ³J_{H,P} = 6.2 Hz).
- 15. Branched trimer 1a was unequivocally characterized by ¹H- & ³¹P-NMR spectroscopy and was found to be identical to our reported preparations described in the ref. 6a.
- 16. ¹H-NMR (90 MHz, CDCl₃ +TMS): 8.2-7.4 (m, 4H, ar); 7.88 (s, 1H, H-8); 6.96 (d, 626.0 Hz, 1H, HP); 6.4-6.2 (m, 1H, H-2'); 5.99 (d, 8.1 Hz, 1H, H-1'); 5.8-5.6 (m, 1H, H-3'); 4.5-4.1 (m, 3H, H-4',5'); 2.7 (m, 6H, CH_2CH_3); 2.15 & 1.99 (2 x s, 6H, Ac); 1.32 (s, 9H, CH₃-tBB); 1.1 (m, 9H, CH₂CH₃). ³IP-NMR (CDCl₃ + ext. standard 85% aq. H₃PO₄): +4.53 (dt, ¹J_{H,P} = 626.0 Hz, ³J_{H,P} = 7.3 Hz).
- ¹H-NMR (270 MHz, CDCl₃ + TMS): 8.30 (s, 1H, H-8); 8.14 (s, 1H, H-2); 8.10 (d, J=8.2 Hz, 1H, H-6); 8.1-6.54 (m, 27H, Ar); 6.27 (d, J=7.3 Hz, 1H, H-1'U); 6.17 (d, 7.7 Hz, 1H, H-1'A); 5.79 (d, 8.2 Hz, 1H, H5); 5.78 (m, 1H, H-3'U); 5.52 (m, 1H, H-2'U); 5.45 (m, 1H, H-2'A); 4.8-4.5 (m, 1H, H-4'A); 4.5-4.4 (m, 3H, H-4' & 5'U); 4.3-4.2 (m, 2H, H-5'A); 2.49 (s, 3H, CH₃Tol); 2.16 & 2.01 (2 x s, 6H, Ac).³¹P-NMR (CDCl₃ + ext. standard 85% aq. H₃PO₄): -1.27 ppm.
- ¹H-NMR (270 MHz, CDCl₃ + TMS): 8.69 (s, 1H, H-8); 8.18 (s, 1H, H-2); 8.03 (d, J=8.2 Hz, 1H, H-6); 8.0-7.2 (m, 14H, Ar); 6.19 (d, J=7.00, 1H, H-1'U); 6.14 (d, 1H, H-1'A); 5.90 (d, 8.2 Hz, 1H, H-5); 5.56-5.47 (m, 1H, H-3'U); 5.42-5.32 (m, 1H, H-2'U); 5.05-4.94 (m, 2H, H-2' & 3'A); 4.74-4.53 (m, 3H, H-4' & 5'A); 4.35-4.19 (m, 3H, H-4' & 5'U); 3.03 (m, 6H, CH₂CH₃); 2.36 (s, 3H, CH₃Tol); 2.07 & 1.95 (2 x s, 6H, Ac); 1.31 (m, 9H, CH₂CH₃). ³¹P-NMR (CDCl₃ + ext. standard 85% aq. H₃PO₄): +0.1 ppm.

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